## Synthesis and Cytotoxic Evaluation of Some 4-Anilinofuro[2,3-b]quinoline Derivatives

by I-Li Chen, Yeh-Long Chen, and Cherng-Chyi Tzeng\*

School of Medicinal and Applied Chemistry, College of Life Science, Kaohsiung Medical University, Kaohsiung City 807, Taiwan

and

## Ih-Sheng Chen

School of Pharmacy, Kaohsiung Medical University, Kaohsiung City 807, Taiwan

Some 4-anilinofuro[2,3-b]quinoline derivatives were synthesized from dictamnine, a natural alkaloid, and evaluated for their cytotoxicity in the NCI's full panel of 60 human cancer cell lines derived from nine cancer cell types, including leukemia, non-small-cell lung cancer, colon cancer, CNS cancer, melanoma, ovarian cancer, renal cancer, prostate cancer, and breast cancer. 1-[4-(Furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone (5) (mean  $GI_{50} = 0.025 \mu M$ ), bearing an 4-acetylanilino substituent at C(4) of furo[2,3-b]quinoline, was more active than its 3-acetylanilino counterpart 7 (mean  $GI<sub>50</sub>=5.27 \mu M$ ), and both clinically used anticancer drugs, N-[4-(acridin-9-ylamino)-3-methoxyphenyl]methanesulfonamide (m-AMSA; mean  $GI_{50} = 0.44 \mu M$ ) and daunomycin (mean  $GI_{50} = 0.044 \mu M$ ). Compound 5 was capable of inhibiting all types of cancer cells tested with a mean  $GI_{50}$ of less than 0.04  $\mu$ M in each case except for the non-small-cell lung cancer (average  $GI_{50} = 1.75 \mu$ M). Although non-small-cell lung cancer is resistant to compound 5, the sensitivity within this type of cancer cells varies: HOP- $62 (G_{50} < 0.01 \,\mu\text{m})$ , NCI-H460  $(G_{50} = 0.01 \,\mu\text{m})$ , and NCI-H522  $(G_{50} < 0.01 \,\mu\text{m})$  are very sensitive, while HOP-92 ( $GI<sub>50</sub> = 12.4 \text{ uM}$ ) is resistant. Among these non-small-cell lung cancers, NCI-H522 was found to be very sensitive to 5, 8a, and 8b with a  $GI_{50}$  values of  $< 0.01$ , 0.074, and  $< 0.01$  µm, respectively.

Introduction. - Acridine derivatives, especially 9-anilinoacridines, have been extensively studied as potential chemotherapeutic agents due to their ability to intercalate in DNA, leading to the inhibition of mammalian topoisomerase II  $[1-5]$ . Among them, N-[4-(acridin-9-ylamino)-3-methoxyphenyl]methanesulfonamide (amsacrine, m-AMSA) has been specifically relevant, and has become a useful clinical drug for the treatment of leukemia and lymphoma [1]. Since then, tremendous effort has been directed toward the design and preparation of new amsacrine analogues with the aim of developing new drug candidates with an improved broad spectrum of antitumor activity  $[6 - 10]$ . Among them, 3-(acridin-9-ylamino)-5-(hydroxymethyl)aniline (AH- $MA$ ) was found to be superior to  $m$ -AMSA against the growth of certain solid tumors such as mammary adenocarcinoma, melanoma, and Lewis lung carcinoma in mice [9]. Unlike  $m$ -AMSA, AHMA, which has a 3,5-disubstituted anilino moiety, was resistant to oxidative metabolism and, therefore, was expected to have longer half-life in plasma [9].

These studies, however, were focused only on the 9-anilinoacridine skeleton, with a wide variety of substituents on the anilino and/or acridine chromophore. No attempt has been carried out concerning the replacement of acridine with its bioisosteric furo[2,3-b] quinoline ring system, which is a constituent of an important group of bioactive natural products such as dictamnine, robustine, and haplopine [11] [12]. The furo[2,3-b]quino-



line system possesses a higher electron density than that of acridine and, therefore, is advantageous, since the major route of breakdown for  $m$ -AMSA in vivo is a nonenzymatically mediated attack of thiol at C(9), resulting eventually in loss of the side chain and the formation of inactive products  $[13-15]$ . Here, we describe the preparation of 3-(furo[2,3-b]quinolin-4-ylamino)-5-(hydroxymethyl)aniline (4;  $c.f.$ Scheme  $1$ ), the bioisosteric isomer of AHMA, in which acridine was replaced with the furo[2,3-b]quinoline moiety. Certain monosubstituted 4-anilinofuro[2,3-b]quinoline derivatives, in which the anilino moiety was substituted with an Ac group and its corresponding oxime and methyloxime at either  $C(3')$  or  $C(4')$  were also prepared. We expect these substituents to form H-bonds with DNA molecule during the intercalation process of the tricyclic furo[2,3-b]quinoline moiety. Recently, we have synthesized certain  $\alpha$ -methylidene- $\gamma$ -butyrolactone-bearing quinolones and evaluated their cytotoxicities on the grounds that, through the intercalation of quinolone, the  $\alpha$ methylidene- $\gamma$ -butyrolactone can specifically alkylate the DNA molecule [16–18]. This versatile  $\alpha$ -methylidene- $\gamma$ -butyrolactone moiety has also been appended onto the 9-anilino group  $(cf. 6)$  in an attempt to prepare a bifunctional compound in which furo[2,3-b]quinoline moiety acts as an intercalator while the lactone ring plays the role of an alkylating unit.

**Results and Discussion.** – The known alkaloid, dictamnine  $(4$ -methoxyfuro $[2,3-b]$ quinoline; 1) was isolated from the root wood of *Zanthoxylum simulans* [11] and was used as the starting material (Scheme 1). Its hydrolysis with HBr in AcOH at 110 $^{\circ}$  for 1 h gave furo[2,3-b]quinolin-4-one (2) in 36% yield [19]. The yield was increased up to 91% by refluxing the reaction mixture for 8 h. This improvement is crucial because the supply of starting dictamnine is limited. Compound  $2$  was chlorinated with POCl<sub>3</sub> to give 4-chlorofuro $[2,3-b]$ quinoline (3)  $[20]$ . Treatment of 3 with 3,5-diaminobenzyl alcohol in a boiling solution of  $EtOH/H<sub>2</sub>O$  2 : 1 gave the desired 3-(furo[2,3-b]quinolin-4-ylamino)-5-(hydroxymethyl)aniline (4) in 64% yield. Accordingly, 1-[4-(furo[2,3-b] quinolin-4-ylamino)phenyl]ethanone  $(5)$  was synthesized from 3 and 4-aminoacetophenone in the boiling EtOH/H<sub>2</sub>O 2 :1 solution. *Reformatsky*-type condensation of 5 with 2-(bromomethyl)acrylate and Zn powder in THF afforded 5-[4-(furo[2,3-b]quinolin-4-ylamino)phenyl]-2,3,4,5-tetrahydro-5-methyl-3-methylidenefuran-2-one (6) in 78% yield.

Preparation of monosubstituted anilino-furo[2,3-b]quinoline derivatives is outlined in *Scheme 2*. Reaction of 5 with NH<sub>2</sub>OH gave exclusively  $(E)$ -1-[4-(furo



 $[2,3-b]$ quinolin-4-ylamino)phenyl]ethanone oxime (8a). The configuration of the oxime moiety was determined by through-space nuclear Overhauser effect spectroscopy (NOESY), which revealed coupling connectivity to Me protons. Accordingly,  $(E)$ -1-[4-(furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone O-methyloxime (8b) was obtained from the reaction of  $5$  and NH<sub>2</sub>OMe. Reaction of  $3$  with 3-aminoacetophenone gave  $1-\left[3-(\frac{\text{fur}}{2},3-b\right]$ quinolin-4-ylamino)phenyl]ethanone (7), which was then treated with either NH<sub>2</sub>OH or NH<sub>2</sub>OMe to afford  $(E)$ -1-[3-(furo[2,3-b]quinolin-4ylamino)phenyl]ethanone oxime (9a) and its Me congener 9b.

All compounds were evaluated in the NCI's full panel of 60 human cancer cell lines derived from nine cancer cell types including leukemia, non-small-cell lung cancer, colon cancer, CNS cancer, melanoma, ovarian cancer, renal cancer, prostate cancer, and breast cancer. For each compound, dose-response curves for each cell line were measured with five different drug concentrations, and the concentrations causing 50% cell-growth inhibition ( $GI<sub>50</sub>$ ) compared with the control were calculated [21] and listed in Table 1. Compound 5 (mean  $GI<sub>50</sub> = 0.025 \mu M$ ), which bears an 4-acetylanilino substituent at  $C(4)$  of furo $[2,3-b]$ quinoline, was more active than its 3-acetylanilino counterpart 7 (mean  $GI<sub>50</sub> = 5.27 \mu M$ ), and both clinically used anticancer drugs,  $m$ -AMSA (mean  $GI_{50} = 0.44 \mu M$ ) and daunomycin (mean  $GI_{50} = 0.044 \mu M$ ). The





Table 1. Inhibition of in Vitro Cancer Cell Lines by Some 4-Anilinofuro[2,3-b]quinoline Derivatives ( $GI<sub>50</sub>$  $[\mu M])^a$ )



a) Data obtained from NCI's in vitro disease-oriented tumor-cell screen.  $GI<sub>50</sub>$ : Drug molar concentration causing 50% cell-growth inhibition. b) Daunomycin.  $\epsilon$ ) Mean values over all cell lines tested. These cell lines are: leukemia (CCRF-CEM, HL-60 (TB ), K-562, MOLT-4, PRMI-8226, and SR ); non-small-cell lung cancer (A549/ATCC, EKVX, HOP-62, HOP-92, NCI-H226, NCI-H23, NCI-H322M, NCI-H460, and NCI-H522); colon cancer (COLC 205, HCC-2998, HCT-116, HCT-15, HT29, KM12, and SW-620); CNS cancer (SF-268, SF-295, SF-539, SNB-19, SNB-75, and U251); melanoma (LOX IMVI, MALME-3M, M14, SK-MEL-2, SK-MEL-28, SK-MEL-5, and UACC-257); ovarian cancer ( IGROV1, OVCAR-3, OVCAR-4, OVCAR-5, OVCAR-8, and SK-OV-3); renal cancer (786-0, A498, ACHN, CAKI-1, RXF 393, SN12C, TK-10, and UO-31); prostate cancer (PC-3 and DU-145); and breast cancer (MCF 7, MCF 7/ADR-RES, MDA-MB-231/ATCC, HS 578T, MDA-MB-435, MDA-N and T-47D ).

cytotoxicity was decreased by converting the Ac group of 5 to the corresponding 1- (hydroxyimino)ethyl 8a (a mean  $GI<sub>50</sub>$  value of 0.35  $\mu$ M) or 1-(methoxyimino)ethyl derivative 8b (0.48  $\mu$ M). The same order was obtained for 3-substituted anilinofuro-[2,3-b]quinoline, in which 1-(hydroxyimino)ethyl and 1-(methoxyimino)ethyl derivatives were less active than their Ac precursor (a mean  $GI<sub>50</sub>$  of 5.27  $\mu$ M for 7, 5.60 for 9a, and 7.56 for **9b**). Compound 4 (mean  $GI_{50} = 31.10 \mu M$ ) was inactive even though its acridine counterpart, AHMA, was proved to be a potent anticancer agent [9]. The present results also show that compound 6 (mean  $GI_{50} = 1.77 \mu M$ ), with both intercalating tricyclic furo[2,3-b]quinoline and alkylating  $\alpha$ -methylidene- $\gamma$ -butyrolactone moieties, was less active than  $5$ ,  $8a$ , and  $8b$ . Compound  $5$  was capable of inhibiting all types of cancer cells tested with a mean  $GI<sub>50</sub>$  of less than 0.04  $\mu$ M in each case except for the non-small-cell lung cancer (average  $GI<sub>50</sub> = 1.75 \mu M$ ). Most of non-small-cell lung cancer cells are also resistant to compounds 8a and 8b. Therefore, the inhibitory activities of 5, 8a, and 8b against the individual non-small-cell lung cancer cells are given in Table 2. Although non-small-cell lung cancer is resistant to compound 5 with an average  $GI<sub>50</sub>$  value of 1.75  $\mu$ m, the sensitivity within this type of cancer cells varies; HOP-62 ( $GI_{50} < 0.01 \mu$ M), NCI-H460 ( $GI_{50} = 0.01 \mu$ M), and NCI-H522 ( $GI_{50} < 0.01 \mu$ M) are very sensitive, while HOP-92 ( $GI_{50} = 12.4 \mu M$ ) is resistant. Among these cancer cells, NCI-H522 was found to be very sensitive to 5, 8a, and 8b with  $GI_{50}$  values of  $0.01, 0.074,$  and  $0.01 \mu$ M, respectively.

Table 2. Growth Inhibition of Non-Small-Cell Lung Cancer Subpanels by 5, 8a, and 8b  $(GI_{50} [\mu M])$ 

						A549/ATCC EKVX HOP-62 HOP-92 NCI-H226 NCI-H23 NCI-H322M NCI-H460 NCI-H522		
$5 \t 0.57$	n.d. <sup>a</sup>	$< 0.01$ 12.4		0.31	0.037	0.68	0.01	< 0.01
$8a \quad 0.45$	4.03	0.18	10.9	11.5	0.38	12.2.	0.21	0.074
$8h \quad 0.72$	3.78	0.35	12.1	3.88	0.57	1 22	0.56	< 0.01
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a) n.d.: Not determined.

**Conclusions.** – Some 4-anilinofuro[2,3-b]quinoline derivatives were synthesized from dictamnine and evaluated for their cytotoxicity in the NCI's full panel of 60 human cancer cell lines. Compound 5, bearing an 4-acetylanilino substituent at C(4) of furo[2,3-b]quinoline, was more active than its 3-acetylanilino counterpart 7, and both clinically used anticancer drugs, m-AMSA and daunomycin. The cytotoxicity was decreased by converting the Ac group of 5 to the corresponding 1-(hydroxyimino)ethyl 8a or 1-(methoxyimino)ethyl derivative 8b. Among nine cancer cell types tested, nonsmall-cell lung cancer is the most resistant to compound 5 with an average  $GI_{50}$  of 1.75  $\mu$ M. However, the sensitivity within this type of cancer cells varies; HOP-62, NCI-H460, and NCI-H522 are very sensitive, while HOP-92 is the most resistant. Further in vivo anticancer evaluation of 5 is ongoing.

## Experimental Part

General. TLC: precoated (0.2 mm) silica gel 60  $F_{254}$  plates from EM Laboratories, Inc.; detection by UV light (254 nm). M.p.: Electrothermal IA9100 digital melting-point apparatus; uncorrected. <sup>1</sup>H-NMR Spectra: Varian Unity-400 spectrometer or Varian Gemini-200 spectrometer, chemical shifts  $\delta$  in ppm with Me<sub>4</sub>Si as an

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internal standard  $(=0$  ppm), coupling constants J in Hz. Elemental analyses were carried out on a *Heraeus*  $CHN-O-Rapid$  elemental analyzer, and results were within  $\pm 0.4\%$  of calc. values.

Furo[2,3-b]quinolin-4(9H)-one (2) [19]. A mixture of 4-methoxyfuro[2,3-b]quinoline (1; 0.20 g, 1 mmol), 48% HBr (10 ml), and AcOH (20 ml) was refluxed for 8 h. After cooling, the mixture was poured into an icecold sat. NaHCO<sub>3</sub> soln. (60 ml), and extracted with AcOEt ( $3 \times 60$  ml). The AcOEt extracts were combined, washed with  $H_2O$ , dried (MgSO<sub>4</sub>), and evaporated to give a residual solid, which was purified by flash column chromatography (FC; silica gel; AcOEt). The proper fractions were combined and evaporated to afford 2  $(0.17 \text{ g}, 91\%)$ . M.p. 236-237°. <sup>1</sup>H-NMR (200 MHz, DMSO): 7.06 ( $d, J = 2.4$ , H $-C(3)$ ); 7.29 ( $m, H-C(6)$ ); 7.56 - 7.68 (m, H - C(2), H - C(7), H - C(8)); 8.28 (d, J = 7.6, H - C(5)). <sup>13</sup>C-NMR (50 MHz, DMSO): 104.85; 105.84; 121.60; 122.12; 123.08; 124.60; 130.06; 140.04; 142.53; 160.70; 166.69.

4-Chlorofuro[2,3-b]quinoline (3) [20]. A mixture of 2 (0.56 g, 3 mmol), POCl<sub>3</sub> (20 ml), and Et<sub>3</sub>N (2 ml) was heated at 110 $^{\circ}$  for 8 h. The mixture was cooled to r.t. and poured into ice-H<sub>2</sub>O (100 ml). NaOH soln. (6N) was added until a pH of 6 resulted. The brown precipitate thus obtained was collected and purified by FC (silica gel; CH<sub>2</sub>Cl<sub>2</sub>) to give **3** (0.45 g, 73%). M.p. 112–113°. <sup>1</sup>H-NMR (200 MHz, DMSO): 6.99 (*d*, *J* = 2.6, H–C(3)); 7.63 (m, H-C(6)); 7.77 (m, H-C(7)); 7.83 (d, J = 2.6, H-C(2)); 8.15 (d, J = 9.8, H-C(8)); 8.32 (m, H-C(5)). <sup>13</sup>C-NMR (50 MHz, DMSO): 104.74; 119.29; 123.82; 124.06; 125.81; 128.80; 129.76; 134.56; 145.26; 147.37; 160.72.

 $3-(Furo/2,3-b/quinolin-4-ylamino)-5-(hydroxymethyl)aniline$  (4). Compound 3 (45 mg, 0.22 mmol) and 3,5-diaminobenzyl alcohol dihydrochloride (47 mg, 0.22 mmol) were dissolved in a boiling soln. of EtOH/H2O 2 : 1 (10 ml). Conc. HCl was added until a pH of 6 resulted, while the reflux was continued for 40 min (TLC monitoring). The solvent was evaporated *in vacuo* to give a residue, to which was added ice-H<sub>2</sub>O (20 ml), and was neutralized with 2N NaOH soln. The resulting precipitate was collected and chromatographed (silica gel;  $CH_2Cl_2/MeOH$  10:1) to give 4 (43 mg, 64%). M.p. 236–237°. <sup>1</sup>H-NMR (400 MHz, DMSO): 4.38 (*d*, *J* = 5.2,  $CH<sub>2</sub>$ ); 5.09 (t, J = 5.2, OH); 5.18 (s, NH<sub>2</sub>); 5.99 (d, J = 2.4, H – C(3)); 6.33 (s, 1 arom. H); 6.40 (s, 1 arom. H); 6.45 (s, 1 arom. H); 7.44 (t,  $J = 8.0$ , H  $-C(6)$ ); 7.67 (t,  $J = 8.0$ , H  $-C(7)$ ); 7.70 (d,  $J = 2.4$ , H  $-C(2)$ ); 7.84 (d,  $J = 8.4$ ,  $H-C(8)$ ); 8.42 (d, J = 8.4, H – C(5)); 9.19 (s, NH). <sup>13</sup>C-NMR (100 MHz, DMSO): 62.89; 102.87; 105.98; 107.48; 108.57; 109.53; 117.16; 122.41; 122.88; 127.90; 128.81; 141.44; 141.68; 143.44; 144.05; 145.53; 149.21; 163.03. Anal. calc. for  $C_{18}H_{15}N_3O_2$   $\cdot$  0.45 H<sub>2</sub>O: C 68.97, H 5.11, N 13.40; found: C 69.48, H 5.10, N 13.03.

 $1-\frac{1}{4}$ -(Furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone (5). From 3 and 4-aminoacetophenone as described for 4. Compound 5 was obtained by FC (silica gel; hexane/AcOEt acetate 2:1) in 77% yield. M.p. 220–221 $^{\circ}$ .  $1H\text{-NMR}$  (200 MHz, CDCl<sub>3</sub>): 2.59 (s, Me); 6.35 (d, J = 2.6, H – C(3)); 7.11 (d, J = 8.8, 2 arom. H); 7.31 (br. s, NH); 7.48 (t, J = 7.4, H - C(6)); 7.57 (d, J = 2.6, H - C(2)); 7.71 (t, J = 8.4, H - C(7)); 7.95 (d, J = 8.4, H - C(5),  $H-C(8)$ ); 8.10 (d, J = 8.8, 2 arom. H). <sup>13</sup>C-NMR (50 MHz, CDCl<sub>3</sub>): 26.34; 105.53; 108.73; 118.04; 119.59; 121.27; 124.37; 128.99; 129.45; 130.20; 131.36; 138.75; 143.92; 145.81; 146.15; 162.82; 196.64. Anal. calc. for C19H14N2O2 : C 75.48, H 4.69, N 9.27; found: C 75.35, H 4.72, N 9.15.

5-[4-(Furo[2,3-b]quinolin-4-ylamino)phenyl]-2,3,4,5-tetrahydro-5-methyl-3-methylidenefuran-2-one (6). To a soln. of 5 (76 mg, 0.25 mmol) in dry THF (10 ml), activated Zn powder (22 mg, 0.32 mmol), hydroquinone (1 mg), and ethyl 2-(bromomethyl)acrylate (65 mg, 0.32 mmol) were added. The mixture was refluxed under  $N_2$ for 2 h (TLC monitoring). After cooling, it was poured into an ice-cold 5% HCl soln. (60 ml) and extracted with  $CH_2Cl_2$  (3 × 60 ml). The combined CH<sub>2</sub>Cl<sub>2</sub> extracts were washed with H<sub>2</sub>O, dried (MgSO<sub>4</sub>), and evaporated to give a residual solid, which was purified by FC (silica gel; hexane/AcOEt 2 : 1). The proper fractions were combined and evaporated to furnish a residual solid, which was recrystallized from AcOEt to afford 6 (72 mg, 78%). M.p. 85 – 87°. <sup>1</sup>H-NMR (400 MHz, DMSO): 1.73 (s, Me); 3.24 (m, CH<sub>2</sub>(4)); 5.79 (s, 1 H, CH<sub>2</sub>=C(3)); 5.88  $(d, J = 2.0, H - C(3'))$ ; 6.12 (s, 1 H, CH<sub>2</sub>=C(3)); 7.29 (d, J = 8.0, 2 arom. H); 7.46 (d, J = 8.0, 2 arom. H); 7.54  $(m, H-C(6))$ ; 7.77  $(m, H-C(2), H-C(7))$ ; 7.92  $(d, J = 8.4, H-C(8))$ ; 8.47  $(d, J = 8.4, H-C(5))$ ; 9.80 (br. s, NH). 13C-NMR (100 MHz, DMSO): 29.12; 41.84; 83.77; 103.66; 105.76; 115.61; 117.35; 122.28; 122.75; 123.21; 123.37; 125.24; 126.47; 129.93; 135.50; 140.26; 140.47; 142.76; 143.70; 161.65; 169.10. Anal. calc. for  $C_{23}H_{18}N_2O_3$ 1.2 H<sub>2</sub>O: C 70.46, H 5.24, N 7.14; found: C 70.51, H 5.31, N 6.91.

 $1-\frac{1}{3}$ -(Furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone (7). From 3 and 3-aminoacetophenone as described for 4. The crude product was recrystallized from AcOEt to give 7 in  $91\%$  yield. M.p.  $158-159^{\circ}$ . <sup>1</sup>H-NMR (400 MHz, DMSO): 2.57 (s, Me); 6.03 (d, J = 2.4, H – C(3)); 7.49 (m, 2 arom. H, H – C(6)); 7.74 (m, 2 arom. H, H – C(7)); 7.82 (d, J = 2.4, H – C(2)); 7.92 (d, J = 8.8, H – C(8)); 8.41 (d, J = 8.4, H – C(5)); 9.58 (s, NH) <sup>13</sup>C-NMR (100 MHz, DMSO): 26.78; 104.72; 105.45; 118.23; 120.38; 122.95; 123.05; 123.16; 125.78; 128.17; 129.17; 129.47; 137.62; 141.50; 142.06; 143.20; 145.57; 162.88; 197.68. Anal. calc. for  $C_{19}H_{14}N_2O_2 \cdot 0.3 H_2O$ : C 74.16, H 4.78, N 9.10; found: C 74.40, H 4.85, N 8.95.

 $(E)$ -1-[4-Furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone Oxime (8a). To a suspension of 5 (61 mg, 0.20 mmol) in EtOH (5 ml) was added NH<sub>2</sub>OH  $\cdot$  HCl (28 mg, 0.40 mmol). The mixture was heated at reflux for 30 min and allowed to cool to r.t. The solvent was removed in vacuo, and the residue was suspended in  $H<sub>2</sub>O$ (20 ml). The precipitate obtained was collected and recrystallized from MeOH to give 8a (63 mg, 99%). M.p. 264 – 265<sup>°</sup>. <sup>1</sup>H-NMR (200 MHz, DMSO): 2.18 (s, Me); 6.09 (d, J = 2.6, H – C(3)); 7.21 (d, J = 8.8, 2 arom. H); 7.49  $(m, H-C(6))$ ; 7.71  $(m, 2 \text{ arom. H}, H-C(7))$ ; 7.81  $(d, J=2.6, H-C(2))$ ; 7.91  $(m, H-C(8))$ ; 8.44  $(d, J=7.6,$ H-C(5)); 9.60 (s, NH); 11.12 (s, NOH). <sup>13</sup>C-NMR (50 MHz, DMSO): 11.39; 104.76; 105.73; 118.24; 121.28; 123.10; 123.21; 126.24; 128.11; 129.13; 131.74; 141.81; 142.11; 142.97; 145.62; 152.40; 162.95. Anal. calc. for  $C_{19}H_{15}N_3O_2 \cdot 0.5 H_2O$ : C 69.98, H 4.94, N 12.88; found: C 70.36, H 4.97, N 12.66.

(E)-1-[4-(Furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone O-Methyloxime (8b). From 5 and 40% NH2O-Me  $\cdot$  HCl as described for 8a: 99% yield: M.p. 167 – 168°. <sup>1</sup>H-NMR (400 MHz, DMSO): 2.19 (s, Me); 3.92 (s, MeO); 6.09 (d, J = 2.8, H – C(3)); 7.19 (d, J = 8.8, 2 arom. H); 7.50 (m, H – C(6)); 7.69 (m, 2 arom. H, H – C(7)); 7.83 (d, J = 2.8, H – C(2)); 7.91 (dd, J = 8.4, 0.8, H – C(8)); 8.38 (d, J = 8.4, H – C(5)); 9.52 (s, <sup>13</sup>C-NMR (100 MHz, DMSO): 12.05; 61.46; 105.12; 105.65; 118.35; 120.81; 123.02; 123.10; 126.57; 128.12; 129.12; 130.26; 141.38; 142.67; 143.14; 145.57; 153.48; 162.85. Anal. calc. for C<sub>20</sub>H<sub>17</sub>N<sub>3</sub>O<sub>2</sub>: C 72.49, H 5.17, N 12.68; found: C 72.33, H 5.24, N 12.53.

(E)-1-[3-(Furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone Oxime  $(9a)$ . From 7 and NH<sub>2</sub>OH HCl as described for 8a: 97% yield. M.p. 248-249°. <sup>1</sup>H-NMR (200 MHz, DMSO): 2.14 (s, Me); 5.97 (d, J = 2.6, H – C(3)); 7.21 (m, 1 arom. H); 7.45 (m, 3 arom. H, H – C(6)); 7.70 (m, H – C(7)); 7.77 (d, J = 2.6, H – C(2)); 7.89  $(dd, J=8.4, 1.0, H-C(8))$ ; 8.42  $(d, J=8.4, H-C(5))$ ; 9.47  $(s, NH)$ ; 11.22 (br. s, NOH). <sup>13</sup>C-NMR (50 MHz, DMSO): 11.46; 103.98; 105.57; 117.87; 119.05; 120.99; 122.46; 122.95; 128.10; 129.10; 137.77; 141.50; 142.22; 142.70; 145.58; 152.51; 162.95. Anal. calc. for C<sub>19</sub>H<sub>15</sub>N<sub>3</sub>O<sub>2</sub> · 0.2 H<sub>2</sub>O: C 71.15, H 4.84, N 13.10; found: C 71.25, H 4.89, N 12.80.

(E)-1-[3-(Furo[2,3-b]quinolin-4-ylamino)phenyl]ethanone O-Methyloxime (9b). From 7 and 40% NH2O-Me  $\cdot$  HCl as described for 8a: 95% yield. M.p. 137 – 138°.  $\cdot$  H-NMR (400 MHz, DMSO): 2.18 (s, Me); 3.90 (s, MeO); 5.97 (d, J = 2.8, H – C(3)); 7.23 (m, 1 arom. H); 7.48 (m, 2 arom. H, H – C(6)); 7.56 (m, 1 arom. H); 7.71 (m, H – C(7)); 7.78 (d, J = 2.8, H – C(2)); 7.90 (d, J = 8.4, H – C(8)); 8.42 (d, J = 8.4, H – C(5)); 9. <sup>13</sup>C-NMR (100 MHz, DMSO): 12.27; 61.59; 104.03; 105.53; 117.88; 119.40; 121.25; 122.86; 122.93; 122.96; 128.09; 129.09; 129.15; 136.76; 141.60; 142.13; 142.75; 145.57; 153.70; 162.96. Anal. calc. for  $C_{20}H_{17}N_2O_2$ : C 72.49, H 5.17, N 12.68; found: C 72.36, H 5.23, N 12.60.

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